

Effect of certain additives on the quality of boar semen during preservation at 15°C and 5°C

T. Saikia, K. Ahmed, P.M. Barua, B.C. Deka and N. Ahmed*

Department of Animal Reproduction, Gynaecology and Obstetrics, College of Veterinary Science, Assam Agricultural University, Assam, India

*Corresponding author: nekibahmeds@gmail.com

Abstract

The study was aimed to evaluate the effect of four additives viz. KMnO_4 , Vitamin E, Butylatedhydroxytoluene and trehalose on quality of boar semen during preservation at 15°C and 5°C up to 96 hours. 24 ejaculates were utilized for the study. Sperm motility was significantly higher with BHT irrespective of preservation temperature and period. However, the mean sperm motility was significantly higher ($P<0.05$) in semen preserved at 15°C than 5°C. Live sperm was significantly ($P<0.05$) lower in Trehalose than KMnO_4 , Vitamin E and BHT. The mean percentage of live intact acrosome irrespective of temperature and preservation period was significantly higher ($P<0.05$) with BHT than others additives. Semen in Modena extender with BHT had significantly higher ($P<0.05$) percentage of HOST-reacted sperm at different preservation periods irrespective of different temperature. In conclusion, Butylatedhydroxytoluene (BHT) was found to be superior to KMnO_4 , Vitamin E and Trehalose for preservation of Hampshire boar semen.

Keywords: Hampshire, semen, additives, preservation, quality

Pig is considered as the most important livestock in North East Region, India, contributes 28% of total country's pig population (Khan *et al.* 2011). Inferior local pigs necessitate the up-gradation with superior germ plasm. Hence, in order to enhance the production potentiality of indigenous pig population of this region, crossbreeding is the only tool with elite exotic breeds through Artificial Insemination (AI). The main idea behind semen preservation is to extend the usefulness of superior germ plasm for the purpose of maximizing number of doses of semen obtainable from a given ejaculate, without

reducing fertility and by extending the fertile life of the doses to facilitate their effectiveness use in breeding (Purdy *et al.* 2008). Several commercial semen extenders have been introduced with the objectives of reducing the metabolic activity during preservation which can be obtained by semen dilution into an appropriate medium and by lowering the temperature (Gadea, 2003). Swine semen cooled at 5°C would be a cheaper alternative to keep with increasing the use of AI. Another benefit of preservation at 5°C is that bacterial growth is reduced, which would improve the quality of semen (Althous and Lu, 2005). However, Reactive oxygen species (ROS) is responsible for sperm dysfunction due to lipid peroxidation of membrane during preservation. To check the level of ROS and promote sperm survival and motility during preservation several additives have been used successfully in supplementation with extender (Roca *et al.* 2005). Keeping in view the facts cited above the study has been planned.

Materials and Methods

Semen samples were collected from four Hampshire boars by simple fist technique. A total of 24 ejaculates were used to evaluate the effect four additives viz. KMnO_4 , Vitamin E, Butylatedhydroxytoluene (BTS) and Trehalose in Modena extender on quality of semen during preservation at 15°C and 5°C up to 96 hours. For preservation, each ejaculate was extended (1:4) with Modena (Weitze, 1991) extender. The extended semen was split into four parts, each part was added with KMnO_4 (6.25 μM), Vitamin-E (100 μM), Butylatedhydroxytoluene (100 μM) and trehalose (2mM) and preserved into 2 ml of micro centrifuge tube at 15°C and 5°C in a BOD incubator up to 96 hours. Prior to evaluation semen was thawed at 37°C for two minutes and gently shaken for homogenization. The evaluation of sperm motility, live sperm, live intact acrosome and HOST reacted sperm at 0 hour (immediately after extension), 24, 48, 72 and 96 hours of preservation.

The data was analyzed statistically by using software SPSS version 17.0.

Results and Discussion

In the present study different additives were utilized to study their effect on quality of boar semen during preservation. The results obtained are presented in Table 1.

Sperm motility

The highest mean percentage of sperm motility was 87.92 \pm 0.79, 76.73 \pm 2.90, 67.08 \pm 2.39, 57.92 \pm 2.96 and 52.00 \pm 3.88 at 0, 24, 48, 72 and 96 hours of preservation

Table 1: Mean sperm motility of Modena extended Hampshire boar semen containing different additives at 15°C and 5°C

Preservation period	Sperm motility									
	15°C					5°C				
	KMNO ₄	Vita-E	BHT	Trehalose	KMNO ₄	Vita-E	BHT	Trehalose	KMNO ₄	Vita-E
0 h	87.33±0.78	87.92±0.79	87.92±0.73	86.67±1.23	87.33±0.78	87.92±0.79	87.92±0.73	86.67±1.23	87.33±0.78	87.92±0.79
24 h	76.67±2.44	72.50±1.90	76.73±2.90	66.25±2.76	55.17±2.88	64.58±3.17	71.67±2.27	54.58±3.82	55.17±2.88	64.58±3.17
48 h	67.08±2.39	60.42±2.21	66.25±3.77	45.83±3.45	52.33±2.23	51.25±2.47	59.33±3.25	35.83±3.48	52.33±2.23	51.25±2.47
72 h	55.00±3.59	52.91±2.59	57.92±2.96	40.00±3.97	48.25±2.97	46.25±2.83	56.25±3.29	25.42±2.57	48.25±2.97	46.25±2.83
96 h	52.00±3.88	45.42±4.03	51.17±4.98	26.25±4.40	42.50±3.76	39.17±3.44	42.08±3.05	20.00±2.57	42.50±3.76	39.17±3.44

Table 2: Mean live sperm of Modena extended Hampshire boar semen containing different additives at 15°C and 5°C

Preservation period	Live sperm									
	15°C					5°C				
	KMNO ₄	Vita-E	BHT	Trehalose	KMNO ₄	Vita-E	BHT	Trehalose	KMNO ₄	Vita-E
0 h	84.33±0.80	75.79±1.68	81.29±1.89	77.96±2.00	84.33±0.80	75.79±1.68	81.29±1.89	77.96±2.00	84.33±0.80	75.79±1.68
24 h	68.08±3.85	73.47±3.37	76.68±1.85	71.01±3.62	66.30±3.00	65.44±2.59	73.42±1.90	60.63±3.76	66.30±3.00	65.44±2.59
48 h	65.50±2.88	68.23±3.78	73.17±2.39	63.00±3.22	62.11±2.72	61.24±2.84	65.43±3.51	55.29±4.08	62.11±2.72	61.24±2.84
72 h	64.02±2.71	67.01±2.68	70.17±2.41	61.08±2.98	61.13±3.13	57.83±3.27	60.04±3.19	53.16±3.32	61.13±3.13	57.83±3.27
96 h	62.15±4.70	57.98±4.25	65.33±4.36	54.58±4.02	56.18±3.29	55.50±4.33	59.68±4.36	51.83±4.45	56.18±3.29	55.50±4.33

at 15°C with Vitamin E, BHT, KMNO₄, BHT and KMNO₄ respectively (Table 1). The corresponding values in semen preserved at 5°C were 87.92±0.79, 71.67±2.27, 59.33±3.25, 56.25±3.29 and 42.50±3.76 with Vitamin E, BHT, BHT, BHT and KMNO₄ respectively.

Analysis of variance revealed that sperm motility differed significantly (P<0.01) among additives, between preservation temperatures and among preservation periods. Results of critical difference test indicated that overall percentage of sperm motility was significantly higher with BHT followed by KMNO₄, Vitamin E and Trehalose irrespective of preservation temperature and period. However, the mean sperm motility was significantly higher (P<0.05) in semen preserved at 15°C than 5°C. There was no available literature on using KMNO₄ as an additive for preservation of semen but beneficial effect of very low concentration of K⁺ on KMNO₄ in cell biology had been confirmed by many workers (Abuladze *et al.* 2009).

In the present study, sperm motility in Modena extender was comparable to the reports of early workers (Funahashi and Sano, 2005; Kadirvel *et al.* 2005). However, the present findings were higher than the reports of Khan *et al.* (2006).

Live sperm

The overall mean percentage of live sperm was significantly (P<0.05) lower in Trehalose than KMNO₄, Vitamin E and BHT, but no significant difference was observed among the later three additives (Table 2). The mean live sperm was significantly higher (P<0.05) in semen preserved at 15°C than 5°C. The mean percentage of live sperm in Modena extender in the current study was higher than the reports of others (Khan *et al.* 2006; Lalrintluanga *et al.* 2009)

Live intact acrosome

The highest percentage of live intact acrosome at 0, 24, 48, 72 and 96 hours of preservation at 15°C was 83.17±0.68, 71.22±2.60, 65.00±2.40, 62.71±2.61 and 52.46±3.95 with KMNO₄, BHT, BHT, BHT and BHT respectively (Table-3). The corresponding values at 5°C were 83.17±0.68, 66.83±2.40, 55.48±3.33, 53.61±3.58 and 49.58±4.24 with KMNO₄, BHT, BHT, BHT and BHT respectively. The overall mean percentage of live intact acrosome irrespective of temperature and preservation period was significantly higher (P<0.05) with BHT than others. The mean live intact acrosome in this study was higher than the report of Boonsorn *et al.* (2010).

Table 3: Mean live intact sperm of Modena extended Hampshire boar semen containing different additives at 15°C and 5°C

Preservation period	Live intact sperm							
	15°C			5°C				
	KMNO ₄	Vita-E	BHT	Trehalose	KMNO ₄	Vita-E	BHT	Trehalose
0 h	83.17±0.68	76.13±1.83	78.13±1.89	76.46±1.91	83.17±0.48	75.13±1.73	76.13±1.49	75.46±1.81
24 h	68.55±3.53	63.47±4.16	71.22±2.60	61.58±3.57	60.51±3.30	55.90±2.58	66.83±2.40	51.25±3.79
48 h	60.08±3.15	59.08±3.52	65.00±2.40	59.56±3.41	51.41±3.51	49.46±3.20	55.48±3.33	43.13±3.90
72 h	53.73±3.36	53.43±3.12	62.71±2.61	46.33±3.21	49.63±3.02	47.88±3.39	53.61±3.58	39.49±3.32
96 h	45.67±4.74	43.25±4.14	52.46±3.95	38.58±4.21	43.17±3.29	42.89±3.10	49.58±4.24	37.83±3.94

Table 4: Mean HOST-reacted sperm of Modena extended Hampshire boar semen containing different additives at 15°C and 5°C

Preservation period	HOST-reacted sperm							
	15°C			5°C				
	KMNO ₄	Vita-E	BHT	Trehalose	KMNO ₄	Vita-E	BHT	Trehalose
0 h	60.50±2.44	61.63±2.21	68.42±2.59	57.92±2.34	59.55±2.44	60.63±2.11	66.42±1.59	56.92±2.74
24 h	53.75±2.21	52.38±2.91	58.92±3.05	49.63±2.94	46.54±2.03	47.54±2.64	56.46±3.03	46.42±2.25
48 h	51.42±3.05	49.92±2.46	55.54±2.50	48.75±2.38	43.42±2.57	46.00±2.26	51.75±2.83	44.83±2.13
72 h	45.58±3.39	47.00±2.55	47.88±2.86	43.63±2.80	41.50±2.98	45.33±2.52	46.75±3.21	42.33±3.75
96 h	41.42±2.79	42.50±2.70	46.25±2.71	41.92±2.64	36.71±2.59	40.58±3.26	42.67±3.31	40.38±3.24

HOST-reacted sperm

Semen in Modena extender with BHT had significantly higher ($P < 0.05$) percentage of HOST-reacted sperm at different preservation periods irrespective of different temperature (Table 4). In the present study, the percentage of HOST-reacted sperm was in close agreement with the findings of Ziaullah *et al.* (2012). However, these findings were higher than that reported by others (Correa *et al.* 2006; Zou and Yang 2000).

Conclusion

The present study concluded that the Butylatedhydroxytoluene (BHT) was found to be superior to $KMNO_4$, Vitamin E and Trehalose for preservation of Hampshire boar semen. This advanced technology could be used to maintain optimum fertility under field condition with superior germplasm.

References

- Abuladze, M.K., Sokhadze, V.M., Namchevadze, E.N., Kiziria, E., Tabatadze, L.V., Lejava, L.V., Gogichashvili, S. and Bakradze, N.B. 2009. Thermal analysis of whole bacterial cells exposed to potassium permanganate using differential scanning calorimetry: A biphasic dose-dependent response to stress. *Sciific. World J.*, **9**: 109-117.
- Althouse, G. and Lu, K. 2005. Bacteriospermia in extended porcine semen. *Theriogenology*, **63**: 573-584.
- Boonsorn, T., Kongbuntad, W., Narkkong, N.A. and Aengwanich, W. 2010. Effects of catechin addition to extender on sperm quality and lipid peroxidation in boar semen. *American-Eurasian J. Agric. Environ. Sci.*, **7**(3): 283-288.
- Correa, M.N., Lucia, T., Bianchi, I., Schmitt, E., Bordignon, J., Rech, D.C., Peruzzo, I.A. and Deschamps, J.C. 2006. Swine semen cooled at 5°C PIGPEL-5 extender: Effect on semen quality in-vitro and fertility estimators in-vivo. *Anim. Reprod.*, **3**(1): 41-48.
- Funahashi, H. and Sano, T. 2005. Selective antioxidants improve the function of extended boar semen stored at 10°. *Theriogenology*, **63**(6): 1605-1616.
- Kadirvel, G., Naskar, S., Das, A. and Hasin, D. 2005. Effect of different extenders on preservation of boar semen at 17°C. *Theriogenology*, **63**: 685-692.
- Khan, M.H., Naskar, S., Das, A. and Bordoloi, R.K. 2006. Comparative efficacy of different diluents on liquid preservation of boar semen. *Indian J. of Anim. Sci.*, **76**(10): 780-783.
- Khan, M.H., Nath, K.C., Deka, B.C., Naskar, S., Bordoloi, R.K., Bhuyan, D. and Sinha, S. 2011. Physical characteristic of Hampshire and crossbred boar semen. *Indian Vet. J.*, **88**(5): 39-41.
- Lalrintluanga, K., Deka, B.C., Borgihain, B.N. and Sarmah, B.C. 2009. Effect of holding time on quality of boar semen preserved at 5°C and 15°C. *Indian Vet J.*, **86**: 1092-1093.
- Purdy, P.H.R., Knox, W., Singleton, K., Spenser, S.F., Spiller, T, Stewart, I., Tharp, N., Welsh, C.S. and Blackburn, H.D. 2008. Preservation on cryopreservation, quality control

and artificial insemination of frozen-thawed boar sperm in National resitory. *Symposium proceeding, Mid West Boar stud Managers Conference St. Louis MO*, **8**: 7-8.

Roca, J., Rodriguez, M.J., Gil, M.A., Carvajal, G., Garcia, E.M., Cuello, C., Vazquez, J.M. and Martinez, E.A. 2005. Survival and in-vitro fertilization of boar spermatozoa frozen in the presence of superoxide dismutase and/ or catalase. *J. Androl.*, **26**(1): 15-24.

Weitz, K.F. 1991. Long term storage of extended boar semen in: Johnson, L. A., 2nd Int. conf. on boar semen preservation, USA. *Reproduction in Domestic Animals*, pp. 231-253.

Ziaullah, M., Ijaz, A., Aleem, M., Mahmood, A.K., Rehman, H., Bhatti, S., Farooq, U. and Sohail, M.U. 2012. Optimum inclusion level of butylatedhydroxytoluene in semen extender improves the quality of post-thawed canine sperm. *Czech J. Anim. Sci.*, **57**(8): 377-381.

Zou, C. and Yang, Z. 2000. Evaluation on sperm quality of freshly ejaculated boar semen in-vitro storage under different temperatures. *Theriogenology*, **53**:1477-1488.

